**STEREOSPECIFIC SYNTHESIS OF DINOSTEROL Arthur Y. L. Shu and Carl Djerassi\* Department of Chemistry, Stanford, University Stanford, California 94305** 

**Abstract: Using a Claisen ortho-ester rearrangement, the biogenetically important marine sterol dlnosterol and its C-24 epimer were synthesized stereospecifically by a sequence which is also attractive for selective isotope labelling in the side chain.** 

**Dinosterol was first isolated from the toxic dinoflagellate, Gonyaulax tamarensis, by**  Shimizu et al<sup>1</sup> and its stereostructure 12 was established unequivocally by X-ray crystallography.<sup>2</sup> Dinosterol (12) and/or its naturally occurring<sup>3</sup>  $\Delta^5$ -3B-hydroxy analog 14 have been proposed<sup>1</sup>,2,4,5 **as possible key precursors in the biosynthesis of the important marine sterol gorgosterol (15).4 In order to verify such proposals, radioactively labelled dinosterol would have to be synthesized. We now report the first synthesis of dinosterol (12) which is equally applicable to the synthesis of 14; most importantly, it lends itself readily to the selective incorporation of isotopic tracers in the side chain.** 

**The starting alcohol 1 (mp 193-194"C, [a]:" +26.6 (CHC13)), obtained by ozonolysis of the tert-butyldimethylsilyl ether of 4a-methyl-5a-dihydrostigmasterol 6 and subsequent NaBH reduction, was oxidized with pyridinium dichromate7 4 to the corresponding aldehyde which was immediately condensed at -78°C with the vinyl lithium reagent prepared from n-BuLi and (E)-2-iodo-2-butene at -60°C. 8 The vinyl iodide in turn was synthesized by the addition of catecholborane to 2**  butyne at 70°C, followed by hydrolysis and treatment of the boranediol with NaOH and I<sub>2</sub>.<sup>9</sup> The **condensation gave two allylic alcohols easily separable on silica gel: the less polar 22R alcohol 2 (73% yield, mp 171-178"C, [a];' +11.9 (CHC13), m/z 516.4354), and the more polar 22s epimer 3 (9% yield, mp 167-17O"C, [a];' +30.5 (CHC13), m/z-516.4306). \_**  <u>m/z</u> 516.4306). The configurational assignment was based upon further conversions (vide infra) of each epimer to the final sterols through the **well-known Claisen rearrangement which transferred the chirality from C-22 to C-24 with con**comitant formation of the **trans-**  $\Delta^{22}$  double bond.<sup>10</sup> The stereochemical assignment is also con**sistent with the fact that condensation of a similar vinyl lithium reagent and aldehyde yielded 11**  as the major product a 22-alcohol with the same absolute configuration as 2.

**Claisen rearrangement of 2 with triethyl orthopropionate <sup>12</sup> and subsequent deprotectlon of the silyl ether with LiBF413 gave a mixture of C-25 epimeric esters, which were separated by reverse phase HPLC (column: Whatman Partisil M9 lo/50 ODS-2; eluent: absolute MeOH): 4a (44% yield**  from 2, mp 159-160°C,  $[a]_D^{20}$  +4.6 (CHCl<sub>3</sub>), m/z 486.4111), and 4b (22% yield, mp 211-214°C,  $[a]_D^{20}$ -7 1 (CHC1<sub>3</sub>), m/z 486.4060). Similar transformation of <u>3</u> and HPLC separation provided 5a (50%

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**yield, mp 163-164°C,**  $\lbrack \alpha \rbrack^{20}_{\text{D}}$  **+11.9 (CHCl<sub>3</sub>), <u>m/z</u> 486.4056); the other C-25 enimer, <u>5b</u>, could not he obtained in pure form.** 

**a Ref. 1 reports mp 220-222°C (CHCl 2 -MeOH), [a] +5(CHCl** ) **for the natural product. A samole of**  dinosterol isolated in our laboratory from the cultured zooxanthellae of the go5gonian **. Briareum asbestinum and purified by HPLC exhlbited mp 212-215°C (hot MeOH), [a], -1 (CHC13).** 

Each of the isomeric hydroxy ester (4a, 4b, 5a) was then subjected to a three-step transformation sequence: protection of the 3B-hydroxy group with tert-butyldimethylsilyl chloride<sup>14</sup> (90% **yield of 6a,6b,7a), lithium aluminum hydride reduction of the C-26 ester function (85% yield of**  8a,8b,9a), mesylation<sup>15</sup> and subsequent reduction with lithium aluminum hydride (93% yield of <u>10</u> and 11). With the resulting destruction of the C-25 epimeric center, the products (10) from 8a and **8b were identical by NMR comparison. Finally deprotection of the silyl ethers gave the free**  sterols (94% yield) 12 and 13. Both exhibited the same GC mobility (3% OV-17, 260°C) as that of natural dinosterol but were separable by reverse phase HPLC (retention times: 12, 1.48; 13, **1.39; cholesterol, 1.00). The physical constants are listed in Table I; unambiguous differentation could be accomplished by 360 MHz NMR comparison (cf. Table** II).

**Table** II. **'H Chemical Shifts of dinosterol (12) and its C-24 epimer 13 (36@ MHz, CDC13/TMS, coupling constants J in Hz) -** 



C-21 was determined by a decoupling experiment involving irradiation of the allylic proton around 2.3 ppm in both isomers.

**<sup>b</sup>around 2.3 ppm in both isomers. C-26 and C-27 were determined by their simultaneous collapse when irradiation occurred around 1.5 ppm.** 



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**By replacing lithium aluminum hydride with the corresponding tritlum or deuterium analog** In one or both of the terminal reduction steps  $(6,7 \rightarrow 8,9 \rightarrow 10,11)$ , appropriate tritium or deuterium **labelling can be effected. Furthermore, by applying the identical reaction sequence to the well-known16 i-methyl ether of the aldehyde 16, the naturally occurring3 23,24-dimethyl-22-dehydrocholesterol (14) and its 24-epimer were also synthesized in our laboratory. -** 

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